STUDIES ON MONOTERPENE GLUCOSIDES—IX^{1, 2} CHEMICAL CORRELATION BETWEEN ASPERULOSIDE AND LOGANIN

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Abstract—The absolute structure of loganin was established by chemical conversion of asperuloside to loganin penta-acetate. The stereochemical relations of a series of the intermediates are also discussed.

LOGANIN^{*}, a bitter glucoside isolated from *Strychnos nux vomica*^{3, 4, 5} and other species of *Strychnos*^{6, 7}, *Menyanthes trifoliata*⁸, Lonicera- and Hydrangea-species⁹, has recently been the subject of various chemical and biosynthetic investigations. Previously, Merz and Birch⁴⁻⁷ showed that this glucoside was a β -glucoside with the formula $C_{17}H_{26}O_{10}$, containing a carbomethoxyl group conjugated with an enol ether, one C-methyl group and a secondary hydroxyl group in its aglycone moiety. Birch further suggested a structural similarity between loganin and plumieride.¹⁰ On the other hand, Büchi¹¹ showed that the acetates **3** and **4** of the two alcoholic C₆-epimers derived from verbenalin (2) differed from loganin pentaacetate. Based on these findings, Ramstad *et al.*¹² assigned the structure **1a** to loganin. This assignment, however, lacks chemical and stereochemical support.

Recently, Battersby¹³ and Arigoni¹⁴ found that loganin was a key intermediate in the biosynthesis of indole alkaloids. However, no information was available on its structure when we started this work.

This paper reports studies on the structure of loganin (1a) including its absolute configuration by chemical transformation of asperuloside $(5)^{15}$ of known absolute structure to loganin pentaacetate.

On catalytic hydrogenation over Pd-C followed by methylation with diazomethane, asperuloside tetraacetate (6) was converted to bisdesoxydeacetylasperulosidic acid methyl ester tetraacetate (7)¹⁶ which was oxidized with osmium tetroxide to the diol 8, $C_{25}H_{34}O_{15}$, m.p. 152–153°. Assuming that the oxidizing reagent approached from the convex side of the aglycone moiety of the compound 7, the newly formed hydroxyl groups in 8 would be β -oriented. The diol 8, after conversion to the pentaacetate 9, $C_{27}H_{36}O_{16}$, m.p. 138–140°, was dehydrated by phosphorous oxychloride and pyridine to a product which gave two spots on a thin layer chromatogram. The nuclear magnetic resonance (NMR) spectrum of this compound shows signals due to vinylic methyl at 8·39 and 8·23 τ which indicate that the dehydrated product is a mixture of two substances 10 and 11. This mixture, without further purification, was subjected to hydrolysis with sodium methoxide followed by acetylation to give ketone 12, $C_{25}H_{32}O_{14}$, m.p. 108°, as the sole crystalline product, derived from 10. The NMR spectrum of 12 showed a signal at 8·85 τ (doublet, J=6 c/s) due to a secondary methyl group and its

^{*} The corresponding free acid, loganic acid, has been isolated from Swertia caroliniensis (C. J. Coscia and R. Guarnaccis, Chem. Commun., 138 (1968)).

ORD curve showed negative cotton effect ($[\phi]_{319} - 7184^\circ$) indicating that the compound 12 was a ketone.

On the other hand, oxidation of 7 with one mole equivalent of *m*-chloroperbenzoic acid gave two isomeric epoxides 13, $C_{25}H_{32}O_{14}$, m.p. 132–134° and 14, $C_{25}H_{32}O_{14}$, m.p. 154°. The NMR spectrum of 13 showed a doublet (J=1 c/s) at 2.62λ due to a vinyl proton on C_3 , a doublet (J=1 c/s) at 6.72τ due to an epoxy proton and a singlet at 8.49τ due to a methyl group on a tertiary carbon. The corresponding signals in the NMR spectrum of 14 appear at 2.75, 6.75 and 8.49τ . These NMR spectra suggest that only the double bond at C_7 – C_8 was oxidized with peracid as is the case with osmium tetroxide, and hence the two oxidation products are stereoisomersdiffering in the configuration of the epoxide ring at C_7 – C_8 . Both 13 and 14, as will be described later, were converted to ketone 12 in fairly good yield.

The configurations of these epoxides were determined by the following reactions.

On treatment with hydrochloric acid in ethylacetate at room temperature, 13 gave chlorohydrin 15, $C_{25}H_{33}O_{14}Cl$, m.p. 171–173°, in fairly good yield. In the same way, 14 gave the corresponding chlorohydrin 16, $C_{25}H_{33}O_{14}Cl$, m.p. 166–167°. The newly formed hydroxyl group in 15 or 16 was resistant to acetylation by the usual method indicating that the chlorine atom occupied the C_7 position.



SCHEME 1

Although nucleophiles are expected to favourably attack the cyclopentane ring from the less hindered β -side, the convex face of the aglyconc moiety, there was practically no difference in the yield of chlorohydrin 15 from 13 and of 16 from 14.

Epoxide 13 was then treated with perchloric acid in acetone to give two diols, 17, $C_{25}H_{34}O_{15}$, m.p. 167–170° and 18, $C_{25}H_{34}O_{15}$, m.p. 91–93° in 14% and 42% yield, respectively. Epoxide 14 was also treated in the same way to afford diols 17 and 18 in yields of 24% and 10%, respectively. On heating with glacial acetic acid, 13 gave pentaacetate 19, $C_{27}H_{36}O_{16}$, m.p. 190–192° in 52% yield, which was identical with the acetate of diol 18. In the same way, epoxide 14 gave pentaacetate 20, $C_{27}H_{36}O_{16}$, m.p. 149–151°, in 27% yield, which was identical with the acetate of diol 17. These results show that C_7 is more accessible than C_8 to the nucleophile, but these results are not sufficient to determine the configuration of the epoxides conclusively.

On the other hand, 13 was treated with BF₃-etherate to give an isomer, $C_{25}H_{32}O_{14}$, m.p. 109°, in 48% yield, which was identified as the ketone 12 derived from 10 by its mixed melting point, and IR and NMR spectra. Ketone 12 was also obtained by analogous treatment of 14, in 41% yield. The fact that both 13 and 14 gave the same compound 12 and that on treatment with sodium methoxide, 12 afforded the deacetylated compound 21, $C_{17}H_{24}O_{10}$, m.p. 193–195°, from which 12 was regenerated by acetylation, indicate that the methyl group at C₈ in 12 has a stable configuration.

Treatment of 14 with BF₃-etherate for a short period, however, afforded a ketone 22, $C_{25}H_{32}O_{14}$, m.p. 146–148°, which was isomerized to ketone 12 by prolonged treatment with the same reagent. This indicates that the C_8 methyl group of compound 22 has an unstable configuration. Since the C_8 methyl group of 12 has a β -orientation, that of ketone 22 should have an α -configuration.

Assuming that this ketone was formed from the epoxides by a backbone hydride rearrangement,¹⁷ 14 should have an α -oriented epoxide ring (Scheme 2). This conclusion was supported by the following NMR spectral data.



SCHEME 2

Compound 14 showed a doublet at $2 \cdot 75 \tau$ (J = 1 c/s) corresponding to the enol proton at C₃ and a doublet at $4 \cdot 40 \tau$ (J = 1 c/s) due to the acetal proton at C₁. The coupling constant of $J_{1,9}$ (1 c/s) is compatible with a conformation in which the C₁ proton is in an equatorial position. In this conformation with an α -oriented epoxide ring, the NMR signal of the C₁ proton, which lies to the side of epoxide ring, is expected to shift to a lower field. On the other hand, the signal of the C₃ proton would shift a little to higher field as this proton is located above the plane of the epoxide ring. On the contrary, neither the C₁ nor C₃ proton of a β -epoxide should be affected by such an anisotropy effect.

In fact, the NMR spectrum of 13, as previously mentioned, shows a doublet (J=1 c/s) assignable to the C₃ proton at 2.62τ and a signal due to the C₁ proton overlapping the methine signals of the glucose moiety at $4.7-5.1 \tau$. Thus comparisons of the NMR signals of 13 and 14 revealed that the epoxide ring in 14 is α -oriented while that in 13 is β -oriented. Consequently, compounds 15-22 should be represented by the absolute structures shown in schemes 1 and 2.

The ketone 12 was reduced with sodium borohydride to yield the 7-hydroxyl compound 23, $C_{23}H_{34}O_{14}$, m.p. 150–151°, $[\alpha]_D - 102 \cdot 7^\circ$, which was converted to the pentaacetate 24, $C_{27}H_{36}O_{15}$, m.p. 144°, $[\alpha]_D - 141 \cdot 8^\circ$. However, as compound 24 was entirely different from authentic loganin pentaacetate, Walden inversion of the 7-hydroxyl group of 23 was attempted.

Treatment of 23 with tosyl chloride gave monotosylate 25, $C_{32}H_{40}O_{16}S$, m.p. 113–115°, which was refluxed with tetraethyl-ammoniun acetate in acetone to yield the epimeric acetate 26, $C_{27}H_{36}O_{15}$, m.p. 132–133°, $[\alpha]_D - 80.4^\circ$. This compound was identified as loganin pentaacetate by comparisons of the IR and NMR spectra, specific rotations and by the mixed melting point. Thus, the absolute configurations of the asymmetric centres at C_1 , C_5 and C_9 of loganin were established as identical with those of asperuloside.



The absolute configuration at C₇ of loganin (1) was determined in the following two ways. First, the benzoate rule¹⁸ was applied to 7-epibenzoylloganin tetraacetate (27) and 7-epiloganin tetraacetate (23). 27, C₃₂H₃₈O₁₅, m.p. 109–110°, $[\alpha]_D = 126 \cdot 5^\circ$, was prepared from 23 by the action of benzoyl chloride in pyridine. The difference in molecular rotation $[M]_{D(27)}$ - $[M]_{D(23)} = -265^\circ$ (in CHCl₃), suggests that the hydroxyl group at C₇ in 23 has an α -configuration. The application of the dissymmetry rule¹⁹ also led to the same conclusion. For, when the molecular rotation of bisdesoxydihydrodea-

cetylasperulosidic acid methyl ester (28)^{16, 20} ($[M]_D - 332^\circ$), in which the C₈ methyl group is already known to be β -oriented, is used as a standard, the predicted shift in rotation caused by the introduction of an α -hydroxyl group into the C₇ position of this compound 28 is -50° (predicted $[M]_D - 382^\circ$), while that due to the β -hydroxyl group is 0° (predicted $[M]_D - 332^\circ$). The observed $[M]_D$ for loganin (1) and 7-epiloganin (29), C₁₇H₂₆O₁₀, derived from 23 are -320° and -392° respectively, in good agreement with the predicted values.

The configuration of the C₈ methyl group was determined as follows. Bisdesoxydihydrodeacetylasperulosidic acid methyl ester tetraacetate (30) was reduced with LAH to give an oily diol 31, which was then acetylated to an oily diacetate 32, $C_{14}H_{22}O_4$. In the NMR spectrum of 32, the two doublets of the terminal methylene protons appear at 4.85 and 5.02 τ . The signal of the two protons due to allylic methylene bearing an acetoxyl group appear at 5.43 τ and another signal of acetoxymethylene protons at 6.10 τ , two acetoxyl groups at 7.90 and 8.00 τ and the C₈ methyl group at 8.95 τ (doublet, J=6c/s). These NMR data support the conclusion that the diacetate can be represented by 32.²¹

In the same way, 7-epitosylloganin tetraacetate (25) was converted to an oily diol and then to its acetate, also an oil. The diacetates derived from 30 and 25 were found to be identical by comparisons of their IR, NMR and mass spectra. Their ORD curves also gave the same sign.

Although we thus tentatively concluded that the C₈ methyl group of loganin (1) is β -oriented, this was still not fully established since identification involved oily substances.

Conclusive proof was obtained as follows.



SCHEME 4

Bisdesoxytetrahydrodeacetylasperulosidic acid methyl ester tetraacetate (33), $C_{25}H_{36}O_{13} \cdot \frac{1}{2}H_2O$, m.p. 154–156°, derived from the corresponding bisdesoxydihydro compound 30 by catalytic hydrogenation over prereduced PtO₂, was reduced with LAH and subsequently acetylated to give pentaacetate 34 as colourless plates, $C_{26}H_{38}O_{13}$, m.p. 98–99°, $[\alpha]_D - 52 \cdot 9^\circ$. The NMR spectrum of 34 shows a signal due to the C₈ methyl group at 9.02 τ (doublet, J=6 c/s) and five acetoxyl groups at 7.94–7.99 τ . On the other hand, 7-epiloganin tetraacetate (23) was hydrogenated in acetic acid with Adams catalyst at 40° to give the dihydro compound 35, $C_{25}H_{36}O_{14}$, m.p. 150–151°, the IR spectrum of which shows no enol band. Reduction of the tosylate 36, $C_{32}H_{42}O_{16}S$, m.p. 145–146°, derived from 35, with LAH followed by acetylation gave colourless plates, m.p. 97–99°, $[\alpha]_D - 45 \cdot 0^\circ$. This compound was identified as 34 by the mixed melting point, $[\alpha]_D$ and by IR and NMR spectral comparisons.

Hence, the absolute configuration of loganin was established to be as in 1.

Recently, Battersby²² and Arigoni²³ also reached the same conclusion in investigations,^{13, 14} by differing methods, on the role of loganin (1) as an intermediate in indole alkaloid biosynthesis. Battersby also found that loganin (1) is a component of *Vinca rosea*.

We recently demonstrated²⁴ that loganin (1) is a precursor of gentiopicroside as well as highly oxygenated iridoid glucosides such as asperuloside.

Addendum—After the preparation of the manuscript of this work we received the report on the structural elucidation of loganin by X-ray analysis of a derivative, which is also in accord with ours (P. J. Lentz, Jun. and M. G. Rossmann, *Chem. Commun.*, 1269 (1969)).

EXPERIMENTAL

M.ps. are uncorrected. NMR spectra were recorded on a Varian A-60 spectrometer. Chemical shifts are given in τ values with TMS as the internal standard and coupling constants (J) in c/s.* Specific rotations were determined with a Rex Photoelectric polarimeter. ORD curves were measured with a Japan Spectroscopic Company ORD/UV-5 spectropolarimeter. Mass spectra were obtained on a Hitachi RMU-6D mass spectrometer equipped with an all glass inlet system and operating with an ionization energy of 80 eV. Silica gel G acc. to Stahl (Merck) was used for thin-layer chromatography. Detection was carried out on exposure to iodine vapour. Silica gel (Mallinckrodt) was used for column chromatography.

Oxidation of Bisdesoxydeacetylasperulosidic acid methyl ester tetraacetate (7) with Osmium tetroxide

A soln of OsO_4 (0.97 g) in ether (10 ml) was added dropwise under stirring to a soln of 7 (2.2 g) in ether (60 ml) containing pyridine (2 ml). After standing the mixture overnight at room temp, the black ppt was collected, washed well with ether and dissolved in EtOH. Gaseous H₂S was passed into the EtOH soln and the resulting ppt was filtered off and washed with EtOH. The washings were combined with the filtrate and evaporated *in vacuo* to give a brown oily residue, which was recrystallized from EtOH to give 0.8 g of the diol 8, m.p. 152–153°. [α]₀¹⁵–90.9° (c=0.29, CHCl₃); IR (Nujol): 3260, 1750, 1640 cm⁻¹; UV (MeOH) λ_{max} : 235 mµ (log ε 3.96); NMR (CDCl₃): 8.82 (3H, s, C_g-Me), 6.29 (3H, s, COOMe), 4.55 (1H, d, J=1.5 c/s, C₁-H) (Found: C, 51.97; H, 6.16. C₂₅H₃₄O₁₅ requires: C, 52.26; H, 5.96%).

Acetylation of cis-Diol 8

8 (600 mg) was dissolved in 2 ml each of pyridine and Ac₂O. After standing overnight at room temp, the reaction mixture was poured into ice water. The resulting solid was recrystallized from EtOH-n-Hexane to yield 506 mg of acetate 9, m.p. 138–140°. $[\alpha]_D^{15} - 121\cdot3°$ (c=0.559, CHCl₃); IR (Nujol): 3450, 1754, 1710, 1640 cm⁻¹; NMR (CDCl₃): 8.75 (3H, s, C_g-Me), 6.30 (3H, s, COOMe), 4.54 (1H, d, J=1 c/s, C₁-H), 2.72 (1H, diffused s, C₃-H) (Found: C, 52.77; H, 5.86. C₂₇H₃₆O₁₆ requires: C, 52.59; H, 5.88%).

* Abbreviations: s = singlet, d = doublet, m = multiplet.

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Epoxidation of Bisdesoxydeacetylasperulosidic acid methyl ester tetraacetate (7)

To a soln of 7 (1.2 g) in benzene (8 ml) was added a soln of *m*-chloroperbenzoic acid (0.45 g) in benzene (8 ml) under cooling and the reaction mixture was allowed to stand overnight in a refrigerator. The soln was washed with 2N NaOH and then with water, dried over MgSO₄ and evaporated *in vacuo* to give a colourless oil (1.13 g), which was chromatographed on silica gel (70 g) using ether as eluent, 5 ml fractions of eluate were collected. The crystalline residue from fractions Nos 32–50 was recrystallized from ether to give colourless needles of epoxide 13, m.p. 132–134° (343 mg). $|\alpha|_D^{15}$ -84·7° (c=0·497, CHCl₃); IR (Nujol): 1740, 1700, 1640 cm⁻¹; UV (MeOH) λ_{max} : 236 mµ (log ε 4·02); NMR (CDCl₃): 8·49 (3H, s, C_g-Me), 6·72 (1H, d, J=1 c/s, C_g-H), 6·29 (3H, s, COOMe), 2·62 (1H, d, J=1 c/s, C_g-H) (Found: C, 54·00; H, 5·65. C₂₃H₂₂O₁₄ requires: C, 53·95; H, 5·80%).

Another epoxide 14 obtained from fractions Nos 61–70, which travelled slower than 13 in TLC (SiO₂-ether), was recrystallized from ether to give colourless needles, m.p. 154° (105 mg). $[\alpha]_D^{20}$ -151·9° (c=0.678, CHCl₃); IR (Nujol): 1740, 1706, 1640, 900 cm⁻¹; NMR (CDCl₃): 8.49 (3H, s, C₄-Me), 6.75 (1H, diffused s, C₄-H), 6.32 (3H, s, COOMe), 4.40 (1H, d, J = 1 c/s, C₄-H), 2.75 (1H, d, J = 1 c/s, C₅-H) (Found: C, 53·92; H, 5·98. C₂₅H₃₂O₁₄ requires: C, 53·95; H, 5·80%).

7-Ketologanin tetraacetate (12)

(a) From pentaacetate 9: POCl₃ (0.6 ml) was added to a soln of 9 (243 mg) in pyridine (2 ml). The reaction mixture was allowed to stand overnight at room temp, and then poured into ice water. The resulting pale brown amorphous substance was dissolved in CHCl₃ and passed through a column of alumina (10 g, Woelm, neutral, activity III). The colourless syrup (195 mg, TLC; SiO₂-ether, 2 spots) obtained on evaporation of the eluate was heated under reflux for 10 min with MeOH-NaOMe. The soln was neutralized on an ion exchange resin (Amberlite IR-120) and the soln was evaporated *in vacuo*. Acetylation of the oily residue in the usual manner followed by chromatography on alumina (15 g, activity III) with ether as eluent furnished the ketone 12, as sole crystalline product. recrystallized from EtOH to give colourless needles. m.p. 108° (65 mg). $[\alpha]_D^{30} - 140^{-7}$ (c = 1·12, CHCl₃); IR (Nujol): 1750, 1740, 1705, 1630 cm⁻¹; NMR (CDCl₃): 8·85 (3H, d, J = 6 c/s, C₈-Me), 6·30 (3H, s, COOMe), 4·57 (1H, d, J = 1 c/s, C₁-H), 2·62 (1H, d, J = 1 c/s, C₃-H) (Found: C, 53·83; H, 5·82. C₂₃H₃₂O₁₄ requires: C, 53·95; H, 5·80%).

(b) From epoxide 13: BF₃-etherate (0.2 ml) was added dropwise to a soln of epoxide 13 (200 mg) in benzene (20 ml) at room temp. The pink coloured reaction mixture was allowed to stand at room temp for 2 h and then poured into ice water. The benzene layer was separated and the aqueous layer re-extracted with benzene. The combined benzene soln was washed with water and dried over MgSo₄. Removal of the solvent gave a crystalline residue, which was recrystallized from EtOH to give fine colourless needles, m.p. 109° (96 mg). This product was identical with 12 obtained from 9. The ketone 12 was also obtained from epoxide 14 by the same treatment (Yield: 83 mg from 200 mg of 14).

Treatment of Epoxide 13 with conc HCl

A soln of 13 (50 mg) in AcOEt (5 ml) containing conc HCl (0.05 ml) was stirred for 15 min to give a colourless syrup which was crystallized from CH_2Cl_2 —light petroleum. Recrystallization from the same solvent afforded fine colourless needles of chlorohydrin 15, m.p. $171-173^{\circ}$ (11 mg), which gave a positive Beilstein flame test. IR (Nujol): 3300, 1745, 1705, 1640 cm⁻¹; NMR (CDCl₃): 8.47 (3H, s, C₄-Me), 6.29 (3H, s, COOMe), 4.62 (1H, d, J=4 c/s, C_1 -H), 2.62 (1H, d, J=1.5 c/s, C_3 -H) (Found: C, 50.50; H, 5.89. $C_{23}H_{33}O_{14}Cl$ requires: C, 50.64; H, 5.61%).

Treatment of Epoxide 14 with conc HCl

To a soln of 14 (50 mg) in AcOEt (5 ml) conc HCl (0.05 ml) was added. After standing for 15 min, the reaction mixture was washed with 10% NaOH and then with water. The AcOEt layer was dried over MgSO₄ and evaporated *in vacuo* to give a colourless oil. Ether was added to the oily residue which crystallized immediately. Recrystallization from CHCl₃-ether gave colourless prisms of chlorohydrin 16, m.p. 166–167° [(38 mg). IR (Nujol): 3300, 1745, 1705, 1640 cm⁻¹; NMR(CDCl₃): 8.47 (3H, s, C_g-Me), 6.29 (3H, s, COO[Me), 4.62 (1H, d, $J = 1 \text{ c/s}, C_1$ —H), 2.62 (1H, d, $J = 1.5 \text{ c/s}, C_3$ —H) (Found: C, 50.37; H, 5.67. C₂₃H₃₃O₁₄Cl requires: C, 50.64; H, 5.61%).

Diol 17 and 18

(a) From epoxide 13: To a soln of 13 (1 g) in acetone (160 ml) was added 6% aq HClO₄ (10 ml). After standing at 38° for 15 h, the reaction mixture was dried over anhydrous K_2CO_3 and evaporated in vacuo.

The resulting brown syrup was chromatographed on silica gel (50 g) using CHCl₃-MeOH (99:1 v/v) as eluent, and 8 ml fractions of eluate were collected. Fractions Nos 87-120 were evaporated to give a crystalline residue which was recrystallized from ether to give 143 mg of 17 as colourless plates, m.p. 167-170°. $[\alpha]_D^{25} - 12.3^\circ$ (c=0.93, CHCl₃); IR (KBr): 3500, 1750, 1696, 1650 cm⁻¹; UV (EtOH) λ_{max} : 237 mµ (log ε 4.02); NMR (CDCl₃): 8.59 (3H, s, C₈-Me), 6.29 (3H, s, COOMe), 4.64 (1H, d, J=4 c/s, C₁-H), 2.65 (1H, d, J=1 c/s, C₃-H) (Found: C, 50.89; H, 6.20. C₂₃H₁₄O₁₅·H₂O requires: C, 50.68; H, 6.11%).

Analogous treatment of fractions Nos 124–235 gave colourless plates of another diol 18, m.p. $91-93^{\circ}$ (423 mg). $[\alpha]_D^{23} - 51 \cdot 4^{\circ}$ (c = 0.63, CHCl₃); IR (KBr): 3550, 3370, 1760, 1700, 1640 cm⁻¹; NMR (CDCl₃): 8.81 (3H, s, C₅-Me), 6.30 (3H, s, COOMe), 4.52 (1H, d, J=2.5 c/s, C₁-H), 2.70 (1H, diffused s, C₃-H) (Found: C, 49.24; H, 6.20. C₂₃H₂₄O₁₅. 2H₂O requires: C, 49.18; H, 6.27%).

(b) From epoxide 14: 14 (500 mg) was dissolved in acetone (80 ml) and 6% aq $HClO_4$ (5 ml) was added. After standing at 38° for 30 h, the soln was neutralized with anhydrous K_2CO_3 and the solvent was removed *in vacuo*. The resulting syrup was chromatographed on silica gel (50 g) with $CHCl_3$ -MeOH (98:2) as eluent and 5 ml fractions of eluate were collected. Fractions Nos 65-70 and 72-84 gave 120 mg of diol 17 and 48 mg of diol 18, respectively.

Pentaacetate 19

(a) From epoxide 13: 13 (500 mg) was dissolved in AcOH (60 ml) and heated on a boiling water bath for 8 h. After cooling, the soln was concentrated *in vacuo* to give a yellow syrup, which was chromatographed on silica gel (40 g) with ether as eluent. Fractions of 5 ml of eluate were collected. Fractions Nos 28–50 gave a crystalline residue which was recrystallized from ether to afford 266 mg of 19 as colourless plates, m.p. 190–192°. $[\alpha]_{D}^{22} - 98.7°$ (c = 0.70, CHCl₃); IR (KBr): 3550, 1760 (shoulder), 1740, 1710, 1650 cm⁻¹; UV (EtOH) λ_{max} : 235 mµ (log ε 4.08); NMR (CDCl₃): 8.80 (3H, s, C₈–Me), 6.29 (3H, s, COOMe), 4.50 (1H, d, J = 2 c/s, C₁–H), 2.69 (overlapped by CHCl₃ signal, C₃–H) (Found: C, 52.33; H, 6.01. C₂₇H₃₆O₁₆ requires: C, 52.59; H, 5.89%).

(b) From diol 18: 18 (80 mg) was dissolved in pyridine (0.8 ml) and Ac_2O (0.8 ml) was added. After standing overnight, the reaction mixture was poured into ice water. The resulting ppt was recrystallized from ether to yield 45 mg of colourless plates, m.p. 190–191°, and identified with compound 19 obtained from 13.

Pentaacetate 20

(a) From epoxide 14: 14 (455 mg) in AcOH (60 ml) was worked up in a similar way to 13 to give 124 mg of 20 as fine colourless needles, m.p. 149–151°. $[\alpha]_D^{22} - 16.9^\circ$ (c=0.76, CHCl₃); IR (KBr): 3500, 1735, 1710, 1645 cm⁻¹; UV (EtOH) λ_{max} : 235 mµ (log ε 4.07); NMR (CDCl₃): 8.69 (3H, s, C_g-Me), 6.29 (3H, s, COOMe), 4.59 (1H, d, J=3.5 c/s, C₁-H), 2.62 (1H, d, J=1 c/s, C₃-H) (Found: C, 52.54; H, 5.89%).

(b) From diol 17: 17 (30 mg) was acetylated in a similar manner to 18 to give 21 mg of colourless needles, m.p. 149-151°. This acetate was identified with compound 20 obtained from 14.

7-Ketologanin (21)

To a soln of 7-ketologanin tetraacetate (12) (300 mg) in hot MeOH (5 ml) was added 0.1N NaOMe soln and the mixture was heated under reflux for 10 min. The soln was cooled and neutralized on an ion exchange resin (Amberlite IR-120), and concentrated *in vacuo* to give a syrupy residue which was crystallized from EtOH-n-Hexane. Recrystallization from the same solvent gave 156 mg of colourless needles, m.p. 193– 195°. NMR (CD₃OD): 8.99 (3H, d, J = 6.5 c/s, C₈-Me), 6.30 (3H, s, COOMe), 4.39 (1H, d, J = 2 c/s, C₁-H), 2.54 (1H, d, J = 1.5 c/s, C₃-H) (Found: C, 51.43; H, 6.27. C₁₇H₂₄O₁₀. $\frac{1}{2}$ H₂O requires: C, 51.41; H, 6.34%). 12 was regenerated by acetylation of this compound 21.

8-Epi-7-ketologanin tetraacetate (22)

A soln of 14 (100 mg) in dry benzene (10 ml) was mixed with BF₃-etherate (0.5 ml), stirred at room temp for 10 min and poured into ice water. The benzene layer was separated and the aqueous layer was reextracted with benzene. The combined benzene soln was washed with water and dried over MgSO₄. Removal of the solvent gave a yellow syrup, which was chromatographed on silica gel (10 g) with CH₂Cl₂ as eluent and 10 ml fractions of eluate were collected. Fractions Nos 22–24 afforded a colourless syrup which was recrystallized from light petroleum-ether to give 42 mg of 22 as colourless needles, m.p. 146–148°. $[\alpha]_{D}^{30} - 76.8^{\circ}$ (c=0.61, CHCl₃); NMR (CDCl₃): 8.84 (3H, d, J=7 c/s, C₂-Me), 6.27 (3H, s, COOMe), 2.54 (1H, diffused s, C₃-H) (Found: C, 53.76; H, 5.87. C₂₂H₃₂O₁₄ requires: C, 53.95; H, 5.80%).

Isomerization of 8-Epi-7-ketologanin tetraacetate (22)

 BF_3 -etherate (0.5 ml) was added to a soln of 22 (26 mg) in benzene (5 ml). The mixture was stirred for 40 min at room temp and then poured into ice water. The benzene layer was separated and dried over MgSO₄. Removal of the solvent afforded a crystalline residue which was recrystallized from ether-light petroleum to give 21 mg of colourless needles, m.p. 109°. This substance was identical with compound 12 in all respects.

7-Epiloganin tetraacetate (23)

To a soln of 12 (44 mg) in dioxane (5 ml) was added NaBH₄ (10 mg) in water (0.5 ml). After standing for 30 min at room temp, AcOH was added, and the mixture was evaporated *in vacuo*. The residue was extracted with CHCl₃, washed with water and dried over MgSO₄. The soln was concentrated *in vacuo* to give an oily residue, which crystallized on addition of ether. Recrystallization from the same solvent gave 23 as colourless needles, m.p. 150–151° (23 mg). $[\alpha]_D^{25} - 102.7^\circ$ (c = 1.08, CHCl₃); IR (Nujol): 3350, 1740, 1690, 1635 cm⁻¹; NMR (CDCl₃): 8.87 (3H, d, J = 6 c/s, C₈-Me), 6.29 (3H, s, COOMe), 2.65 (1H, diffused s, C₃-H) (Found: C, 53.68; H, 6.01. C₂₃H₃₄O₁₄ requires: C, 53.76; H, 6.13%).

7-Epiloganin pentaacetate (24)

23 (50 mg) was acetylated with 1 ml each of pyridine and Ac₂O to give 18·2 mg of **24** as colourless needles on crystallization from EtOH, m.p. 144°. $[\alpha]_{3^0}^{3^0} - 141\cdot8^\circ$ (c = 0.71, CHCl₃); IR (Nujol): 1753, 1708, 1635 cm⁻¹; NMR (CDCl₃): 8·90 (3H, d, J = 6 c/s, C₈-Me), 6·30 (3H, s, COOMe), 2·70 (1H, d, J = 1 c/s, C₃-H) (Found: C, 53.95; H, 5·99. C₂₇H₃₆O₁₅ requires: C, 54.00; H, 6.04%).

7-Epitosylloganin tetraacetate (25)

Tosyl chloride (300 mg) was added to a soln of 23 (100 mg) in pyridine (1.5 ml) and the mixture was allowed to stand at room temp for 12 h. Then it was poured into ice water and the resulting ppt was recrystallized from EtOH to give 84 mg of tosylate 25 as colourless needles, m.p. $113-115^{\circ}$. $[\alpha]_{D}^{20}-95\cdot4^{\circ}$ (c = 0.89, CHCl₃); NMR (CDCl₃): 9.00 (3H, d, J=6 c/s, C₈-Me), 7.55 (3H, s, aromatic Me), 6.32 (3H, s, COOMe), 2.65 (3H, diffused d, J=8 c/s, C₃-H and aromatic H), 2.25 (2H, diffused d, J=8 c/s, aromatic H) (Found: C, 52.64; H, 5.91. C₃₂H₄₀O₁₈S.H₂O requires: C, 52.60; H, 5.80%).

Epimerization of 7-Epitosylloganin tetraacetate (25)

A soln of 25 (73 mg) and tetraethylammonium acetate (164 mg) in acetone (3 ml) was heated under reflux for 22 h and then the solvent removed *in vacuo*. The residue was dissolved in CHCl₃ and washed with water and sald ag NaCl. The CHCl₃ soln was dried over MgSo₄ and evaporated to give a brown oily residue which was chromatographed on silica gel (30 g) using CHCl₃ as eluent and 2 ml fractions of eluate were collected. Fractions Nos 25-39 were combined and evaporated to give a crystalline residue. Recrystallization from AcOEt-light petroleum afforded 20 mg of colourless needles, m.p. 132-133°. $[\alpha]_D^{25} - 80.4°$ (c = 0.77, CHCl₃); IR (Nujol): 1760, 1738, 1710, 1645 cm⁻¹; NMR (CDCl₃): 8.99 (3H, d, J = 6 c/s, C₈-Me), 6.32 (3H, s, COOMe), 2.72 (1H, D, J = 1 c/s, C₃-H) (Found: C, 53.93; H, 6.17. C₂₇H₃₆O₁₅ requires: C, 54.00; H, 6.04%). This compound was identical with authentic loganin pentaacetate in all respects.

7-Epibenzoylloganin tetraacetate (27)

A mixture of 23 (54 mg), C_eH_3COCI (40 mg) and pyridine (2 ml) was allowed to stand overnight at room temp, and then poured into ice water. The resulting ppt was recrystallized from ether-light petroleum to give colourless needles, m.p. 109–110° (37 mg). $[\alpha]_D^{25} - 126.5°$ (c =0.65, CHCl₃); IR (Nujol): 1745, 1710, 1625, 1600 cm⁻¹; NMR (CDCl₃): 8.79 (3H, d, J = 6 c/s, C_g -Me), 6.32 (3H, s, COOMe), 2.75–1.90 (6H, C_3 -H and aromatic H) (Found: C, 57.69; H, 6.00. $C_{32}H_{38}O_{15}$ requires: C, 57.99; H, 5.78%).

7-Epiloganin (29)

To a soln of 23 (130 mg) in MeOH (3 ml) was added 0.1N NaOMe (0.1 ml) and the mixture was heated under reflux for 15 min. After cooling, the soln was neutralized on an ion exchange resin (Amberlite IR-120). The filtrate was evaporated *in vacuo* to give a pale yellow syrup, which was chromatographed on a Carbon column consisting of 5 g each of carbon and celite using water as eluent. Evaporation of the eluate yielded a colourless syrup. $[\alpha]_{D}^{30} - 100.5^{\circ}$ (c = 0.64, EtOH); NMR (D₂O): 8.99 (3H, d, J = 6 c/s, C₃--Me), 6.29 (3H, s, COOMe), 3.75 (1H, d, J = 3.5 c/s, C₁-H), 2.54 (1H, d, J = 1 c/s, C₃-H) (Found: C, 49.44; H, 6.91. C₁₇H₂₆O₁₀·H₂O requires: C, 49.67; H, 6.91%).

Reduction of Bisdesoxydihydrodeacetylasperulosidic acid methyl ester tetraacetate (30) with LAH

A soln of **30** (2·4 g) in THF (20 ml) was added dropwise to a suspension of LAH (3 g) in ether (80 ml). The reaction mixture was refluxed for 3 h. After decomposition of excess reagent with AcOEt, 2N NaOH soln was added to the mixture to dissolve the inorganic salts. The soln was extracted with ether and dried over MgSO₄ and the solvent removed. The resulting colourless syrup (360 mg) was then applied to an alumina column (10 g), eluted first with benzene (50 ml) and then with benzene-ether (6:4 v/v; 100 ml). Fractions with the latter eluent provided 147 mg of a colourless syrup **31**. IR (CHCl₃): 3350, 1640, 905 cm⁻¹; NMR (CDCl₃): 4·80, 5·07 (2H, C=CH₂), 5·89 (2H, s, allylic CH₂OH), 6·50 (2H, m, CH-CH₂OH). This substance **31** was acetylated in the usual manner to give a colourless syrup which was chromatographed on silica gel (10 g). The column was eluted successively with benzene (40 ml), benzene-ether (99:1 v/v; 80 ml) and finally benzene-ether (98:2 v/v; 80 ml); 4 ml fractions were collected. Fractions Nos 37–48 (benzene-ether 98:2 v/v) yielded 58 mg of diacetate **32** as a colourless syrup. IR (CHCl₃): 1730, 1650, 900 cm⁻¹; NMR (CDCl₃): 4·85, 5·02 (2H, C = CH₂), 5·43 (2H, s, allylic CH₂OAc), 6·10 (2H, m, CH—CH₂OAC), 7·90, 8·00 (2×OAc), 8·95 (3H, d, J=6 c/s, CH—CH₃) (Found: C, 66·19; H, 8·89. C₁₄H₂₂O₄ requires: C, 66·13; H, 8·72%).

Reduction of 7-Epitosylloganin tetraacetate (25) with LAH

A soln of 25 $(1\cdot12 \text{ g})$ in THF (20 ml) was added dropwise to a suspension of LAH (2 g) in ether (60 ml). The reaction mixture was worked up in an analogous way to 30 to give a yellow syrup (140 mg), which was chromatographed on alumina (8 g) with benzene and then benzene-ether (1:1 v/v) as eluents. The eluate with benzene-ether (1:1 v/v) afforded a syrup of the diol 31 (21 mg). This substance was acetylated in the usual manner and chromatographed on alumina (5 g) using ether as eluent to furnish a colourless syrup of the diacetate 32 (9 mg). This was identified with the acetate 32 obtained from 30 by comparisons of the IR, NMR and mass spectra.

Bisdesoxytetrahydrodeacetylasperulosidic acid methyl ester tetraacetate (33)

30 (320 mg) was hydrogenated in AcOH (30 ml) with prereduced PtO_2 (200 mg) at atmospheric pressure for 3 h. After removal of the catalyst by filtration, the filtrate was evaporated *in vacuo* to give a crystalline residue. Recrystallization from EtOH afforded a colourless prisms of the tetrahydro compound **33**, m.p. 154–156° (106 mg). $[\alpha]_{D}^{25}-61\cdot0°$ (c=0.42, CHCl₃); IR (Nujol): 1760 cm⁻¹; NMR (CDCl₃): 8-97 (3H, d, J=5 c/s, C_g-Me), 6.35 (3H, s, COOMe) (Found: C, 54·24; H, 6·73. C₂₅H₃₆O₁₃, $\frac{1}{2}$ H₂O requires: C, 54·18; H, 6·81%).

Reduction of Bisdesoxytetrahydrodeacetylasperulosidic acid methyl ester tetraacetate (33) with LAH

A soln of 33 (259 mg) in THF (20 ml) was added to a suspension of LAH (280 mg) in ether (20 ml) with stirring and the mixture was refluxed for 4 h. After decomposition of the excess reagent with AcOEt, satd aq Na₂SO₄ was added to the reaction mixture and a white ppt formed. The soln was decanted off and the ppt was triturated with EtOH. The ethanolic soln was neutralized on an ion exchange resin (Amberlite IR-120) and evaporated *in vacuo*. The resulting oily residue was acetylated with pyridine and Ac₂O in the usual manner to give a crystalline substance. Recrystallization from EtOH-n-hexane afforded 94 mg of pentaacetate 34 as colourless prisms, m.p. 98–99°. $[\alpha]_{2^6}^{2^6} - 52.9^\circ$ (c = 0.97, CHCl₃); IR (Nujol): 1740, 1240–1210 cm⁻¹; NMR (CDCl₃): 9.02 (3H, d, J = 6 c/s, C_8 -Me), 7.94–7.99 (15H, 5 × OAc), 6.30 (5H, m, C_3 , C_{11} and C_5 -H) (Found: C, 55.67; H, 6.61. $C_{26}H_{38}O_{13}$ requires: C, 55.91; H, 6.85%).

Dihydro-7-epiloganin tetraacetate (35)

A soln of 23 (262 mg) in AcOH (10 ml) was shaken with H_2 at 40° for 22 h in the presence of a catalyst prepared by the reduction of 180 mg of PtO₂. The catalyst was filtered off and the filtrate was evaporated to dryness. The resulting colourless syrup was crystallized from EtOH-n-hexane to afford 235 mg of 35 as fine colourless needles, m.p. 150–151°. $[\alpha]_{D}^{10}$ – 50.6° (c =0.77, CHCl₃); IR (Nujol): 3400–3150, 1740; NMR (CDCl₃): 8.89 (3H, d, J=6 c/s, C₈–Me), 6.34 (3H, s, COOMe), 6.21 (2H, m, C₃–H) (Found: C, 53.85; 6.55. C₂₃H₃₆O₁₄ requires: C, 53.56; H, 6.48%).

Dihydro-7-epitosylloganin tetraacetate (36)

A soln of 35 (276 mg) and tosyl chloride (360 mg) in pyridine (3 ml) was stood overnight at room temp and then poured into ice water. The resulting crystalline substance was collected and recrystallized from EtOH to give 194 mg of fine colourless needles, m.p. 145–146°. $[\alpha]_{p^0}^{30}$ –60.0° c=0.60, CHCl₃); NMR (CDCl₃): 9.05 (3H, d, J=7 c/s, C_g-Me), 7.55 (3H, s, aromatic Me), 6.35 (3H, s, COOMe), 2.64, 2.22 (4H, AB quartet, J=8 c/s, aromatic H) (Found: C, 53.61; H, 6.23. C₃₂H₄₂O₁₀S requires: C, 53.78; H, 5.93%).

Reduction of Dihydro-7-epitosylloganin tetraacetate (36) with LAH

A soln of 36(190 mg) in THF (5 ml) was added dropwise to a suspension of LAH (250 mg) in ether (10 ml) with stirring and the mixture was heated under reflux for 4 h. After decomposition of the excess reagent with AcOEt, satd aq Na₂SO₄ was added to the reaction mixture and a white ppt formed. The soln was decanted off and the residue was triturated repeatedly with EtOH. The ethanolic soln was neutralized on Amberlite IR-120 and evaporated *in vacuo* to give colourless syrup which was acetylated in the usual way. The resulting pale yellow syrup was chromatographed on silica gel (5 g) eluting first with light petroleum (4 ml) and then light petroleum-ether (1:1 v/v; 10 ml) and 1 ml fractions of eluate were collected. Fractions Nos 7–9 gave a colourless prisms, m.p. 97–99°. $[\alpha]_D^{25}$ – 45.0° (c=0.39, CHCl₃) (Found: C, 56.13; H, 7.10. C₂₄H₁₄O₁₃ requires: C, 55.91; H, 6.85%).

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